

MULTISCREEN™ TR-FRET cAMP 1.0 No-Wash Assay Kit

INTRODUCTION

Multiscreen™ TR-FRET cAMP 1.0 No Wash Assay Kits provide a homogenous TR-FRET assay method for adenylyl cyclase activity detection in 96-well, 384-well and higher multiplexity. α -cAMP antibody (Ab) is labeled with MultiScreen™Eu while cAMP is labeled with MultiScreen™650. In the absence of cAMP, MultiScreen™650-cAMP is bound to MultiScreen™Eu- α -cAMP-Ab to give a strong TR-FRET Emission at 655 nm. Free cAMP in the test sample competes for binding to the MultiScreen™ Eu- α -cAMP-Ab, reducing TR-FRET signal from MultiScreen™650-cAMP binding. The MultiScreen™650-cAMP only has fluorescence lifetime of nanosecond while MultiScreen™Eu- α -cAMP-Ab has much longer fluorescence lifetime value due to TR-FRET. The magnitude of TR-FRET is proportional to the concentration of cAMP in a sample. Multiscreen™ TR-FRET cAMP 1.0 No Wash Assay Kits are validated by Multiscreen™ stable cell lines.

| Kit Components | Catalog Numbers | | | Storage | Instrument Platform |
|---|--|-----------------------------------|-------------------------------------|----------------------|----------------------------|
| | MSCM01-1 (0.5K tests, 384-well) | MSCM01-10 (5K tests, 384-well) | MSCM01-100 (25K tests, 384-well) | | |
| Component A: MultiScreen™Eu- α -cAMP-Ab | 1vial | 1 vial 50x | 5 vials 50x | -4°C and avoid light | TR-FRET microplate readers |
| Component B: MultiScreen™650-cAMP | 1vial | 1 vial 50x | 5 vials 50x | | |
| Component C: cAMP Standard | 1 vial (20 μ L 1mM) | 1 vial (33 μ g) | 1 vial (33 μ g) | | |
| Component D: Cell Lysis Buffer | 10 mL | 100 mL | 100 mL 5 bottles | | |
| Component E: Diluent | 10 mL | 100 mL | 100 mL 5 bottles | | |
| Reagents NOT included in the kit | Assay Buffer: Hank's Balanced Salt Solution with 20mM HEPES, pH 7.4; When necessary: IBMX (3-Isobutyl-1-methylxanthine); Forskolin (Adenylyl cyclase activator) | | | | |

PREPARE WORKING SOLUTIONS (Mix well by gentle-vortexing or pipette mixing after each step):

- cAMP standard:** Add 100 μ L Component E to Component C to make 1mM stock solution for MSCM01-10 and MSCM01-100 kits only and gently vortex to mix: Add 1 μ L 1mM stock solution into 99 μ L Component E or cell culture media to make 10 μ M standard followed 4-fold serial dilution to make 10000, 2500, 625, 156.3, 39.1, 9.8, 2.4, 0.61, 0.15, 0.038 nM final concentrations. Mix gently with a pipette after each dilution. Add 10 μ L or 20 μ L of serial diluted cAMP standard per 384-well in microtiter assay plate after last the incubation period from step 5.
- MultiScreen™Eu- α -cAMP-Ab working solution:** Add 50 μ L Component A to 2.5mL Component D (scale down based on need). Prepare right before use. Store at 4°C.
- MultiScreen™650-cAMP working solution:** Add 50 μ L Component B to 2.5mL Component D (scale down based on need). Prepare right before use. Store at 4°C.

cAMP ASSAY PROTOCOL (384-well format)

- Prepare cells** (Evaluate each cell line to determine optimal cell density and other conditions.)
Adherent: Plate cells overnight in growth media with 10% FBS at 3k-9k cells/40 μ L/384-well in Poly-D-Lysine coated white opaque bottom plate. Remove growth media carefully before compound treatment.
Suspension: Centrifuge the cells from the culture media and then suspend the cell pellet in the appropriate amount of Assay Buffer at 3k-12k cells/5 μ L/ 384-well in small volume, white, opaque bottom plate.
- Compound Treatment** (The incubation time and temperature can be optimized for each receptor): Add 15 μ L of test compounds to adherent cells or 5 μ L to suspension cells per well and incubate for 20 minutes at 37°C.
- Termination (30 μ L or 20 μ L final volume):** Add 7.5 μ L MultiScreen™650-cAMP working solution and 7.5 μ L MultiScreen™Eu- α -cAMP-Ab working solution sequentially to adherent cells per well or 5 μ L of each sequentially to suspension cells per well. Incubate 30 minutes at room temperature in the dark. Read fluorescence emission on a TR-FRET compatible reader at 665 nm and 620 nm.

